



**Original article**

**Alloimmunization in Patients with Sickle Cell Disease and Thalassemia: Experience of a Single Centre in Oman**

Salam Alkindi,<sup>1,2</sup> Saba AlMahrooqi,<sup>2</sup> Sumaiya AlHinai,<sup>2</sup> Ali AlMarhoobi,<sup>1,2</sup> Saif Al-Hosni,<sup>2</sup> Shahina Daar,<sup>1,2</sup> Naglaa Fawaz<sup>2</sup> and Anil Pathare<sup>2</sup>

<sup>1</sup> College of Medicine & Health Sciences, Sultan Qaboos University, Muscat, Oman.

<sup>2</sup> Department of Haematology, Sultan Qaboos University Hospital, Muscat, Oman.

**Competing interests:** The authors have declared that no competing interests exist.

**Abstract. Background:** Blood transfusion is an integral part of the supportive care for patients with sickle cell disease (SCD) and thalassaemia. The hazard of red cell alloimmunization, however, is one of the main complications of this therapy.

**Objectives:** The aim of this study was to evaluate the prevalence of red cell alloimmunization in Omani patients with sickle cell anaemia and thalassemia.

**Methods:** This study included 262 patients whose historical transfusion records were available. One hundred and twenty-nine patients with thalassaemia who were attending the day care unit for regular transfusions, and 133 SCD patients admitted at our hospital were included in this study. The Diamed® gel system was used for the screening and identification of atypical antibodies.

**Results:** The rate of alloimmunization in SCD patients was 31.6% (n=42, 95%CI, 24.87-40.66), whereas in patients with thalassaemia it was 20% (n=26; 95%CI, 13.9-27.6). Antibodies to E, e, C, c, D, K, S, Fy<sup>a</sup>, Kp<sup>a</sup>, Jk<sup>a</sup> and C<sup>w</sup> were observed; 85% of the patients were also immunised with Rh and Kell antigens. Considering the two groups together, 8 developed nonspecific antibodies and 12 developed more than one antibody.

**Conclusions:** Red cell transfusions were associated with a significant risk of alloimmunization. It is, therefore, imperative to perform an initial extended red cell phenotyping for both donors and recipients, and carefully select ABO, Rh and Kell matched donors. The higher incidence of alloimmunization in SCD patients is related to the inherent SCD-specific inflammatory state.

**Keywords:** Multitransfused; Alloimmunization; Antibodies; Blood Transfusion; SCD; Thalassaemia.

**Citation:** Alkindi S., AlMahrooqi S., AlHinai S., AlMarhoobi A., Al-Hosni S., Daar S., Fawaz N., Pathare A. Alloimmunization in patients with sickle cell disease and thalassemia: experience of a single centre in Oman. *Mediterr J Hematol Infect Dis* 2017, 9(1): e2017013, DOI: <http://dx.doi.org/10.4084/MJHD.2017.013>

**Published:** February 15, 2017

**Received:** October 9, 2016

**Accepted:** January 9, 2017

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by-nc/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Correspondence to: Dr. Salam Alkindi, BA, MB, BCh, BAO, DME, MSc, FRCP. Consultant Haematologist, Department of Haematology, Professor, College of Medicine & Health Sciences, Sultan Qaboos University, P. O. Box 35, Muscat 123, Sultanate of Oman. Tel: +96-824411182, Fax: +96-824413419. E-mail: [sskindi@yahoo.com](mailto:sskindi@yahoo.com)

**Introduction.** Sickle cell disease and thalassaemia are the most frequent genetic disorders in Oman with a combined carrier frequency rate of about 6%.<sup>1-3</sup> Furthermore, in these congenital haemolytic disorders, there are limited curative options. Thus,

long-term blood transfusion remains an integral treatment option for these conditions, in order not only to save life but more importantly to improve the quality of life.<sup>4</sup>

Development of anti-RBC antibodies (alloantibodies and autoantibodies) can significantly complicate transfusion therapy.<sup>5-7</sup> Furthermore, some of these alloantibodies being haemolytic, can cause haemolytic transfusion reactions, and thereby limit the utility of further transfusion, whereas others are clinically insignificant.<sup>8</sup> Erythrocyte autoantibodies appear less frequently, but they can result in clinical hemolysis and difficulty in cross-matching compatible blood units.<sup>9</sup> Patients with autoantibodies may have a higher transfusion rate and often require immunosuppressive drugs, splenectomy or alternative treatments to maintain an adequate level of haemoglobin.

Despite the recognition of antibodies as a transfusion-associated risk,<sup>7,10-13</sup> little is known about the extent and causes of these phenomena among thalassaemia and sickle cell disease patients from the Sultanate of Oman or the most appropriate methods of prevention. Approaches for prevention or treatment of alloimmunizations are under debate and include the provision of RBCs matched for all the major antigens associated with clinically significant antibodies, or to only give blood matched for antibodies that have already been detected. The reason for such a controversy may lie in the fact that many alloantibodies are not harmful and that expensive prevention methods may, therefore, benefit only some patients.<sup>14</sup> In addition, donor feasibility and the cost of RBC matching could impact on these approaches as also the own local guidelines regarding this issue. Furthermore, a better knowledge basis of the potential harmful antibodies among the thalassaemia and sickle cell disease patients can assist in considering the appropriate transfusion strategy to use. Our objective was to assess the prevalence of alloimmunization among our multiply transfused patients with thalassaemia and sickle cell anaemia.

**Materials and Methods.** Diagnosis of homozygous thalassaemia major and sickle cell disease was initially made by high-performance liquid chromatography [HPLC] profiles. However, it was further confirmed with family member studies [parents] and where necessary, by DNA studies using Sanger sequencing.

*Thalassaemia patients:* Clinical features and transfusion records of 129 thalassaemia patients,

aged 5-32 years, 44 males, 85 females, who received regular transfusion were analysed. These patients were attending the day care unit at SQUH for regular transfusions.

*Sickle cell anaemia patients:* 133 sickle cell disease patients [113 SS and 20 S-beta thal] who were admitted to SQUH haematology wards (30 males and 103 females) and who received regular transfusion were analysed. The transfusion records of all the patients including those transfused for their first time were examined for the presence of alloimmunization and antibody specificity, age, gender and ethnicity.

*Donors:* Blood donors from the SQUH blood bank were identified for their racial background, and RBC phenotype was performed for the following antigens C, c, D, E, e and Kell. The donor's ethnic origin was classified into Arabs and non-Arabs.[Data not shown]

*Laboratory protocol. Antibody screening:* Detection of alloantibodies was performed on a fresh blood sample using the indirect antiglobulin test by the column agglutination method. The gel card centrifugation technique was used (DiaMed AG, Cressier sur Morat, Switzerland). All patients were screened before any transfusion.

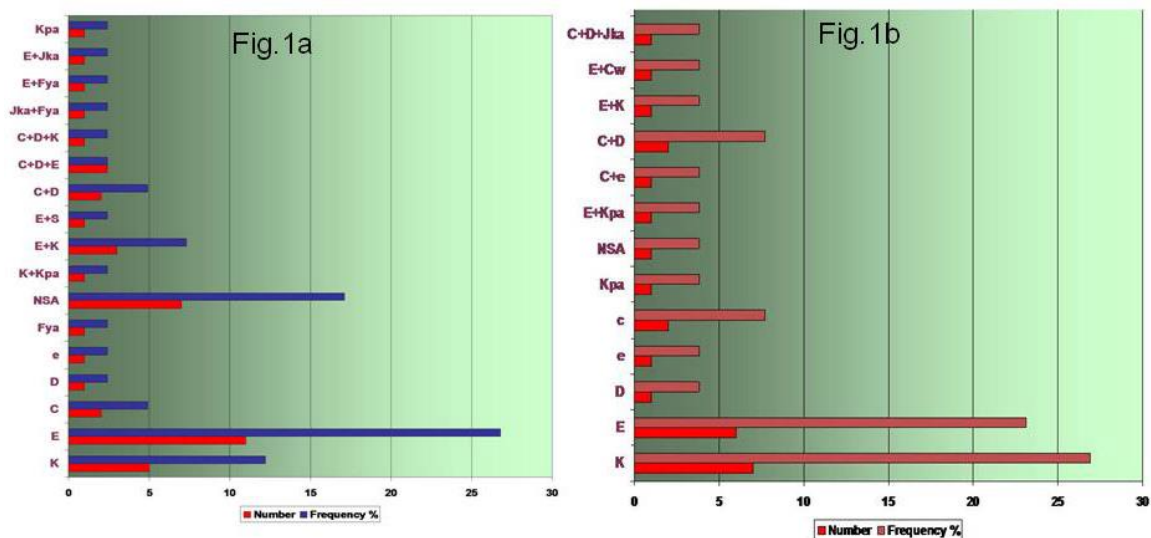
*Antibody identification:* Antibody specificity was determined using a standard panel of red cells reacting to known antigens using column agglutination and gel centrifugation (ID-DiaPanel and ID-DiaPanel-P, DiaMed AG). The indirect antiglobulin test and enzymatic papain-treated RBC test at 37°C were performed when necessary and elution of antibodies was done to help in identification. Detection of alloantibodies masked by autoantibodies required the use of adsorption techniques using Polyethylene glycol [PG], or albumin or low-ionic strength saline [LISS] to identify the underlying antibody by the indirect antibody test [IAT].

**Results.** *Thalassaemia:* 26(20%) of the 129 patients had positive antibody screening, in whom 34 IgG alloantibodies were detected (95% CI, 13.9-27.6). 18(69%) patients developed one antibody; 6(23%) developed two antibodies and one (4%) developed three antibodies. One of the patients presented with a non-specific antibody (NSA). **Table 1** and **Figure 1b** shows the specificities and

**Table 1.** Type and frequency of antibodies identified in Thalassemia and SCD patients

Disease	Thalassemia		Sickle cell disease	
	No. of patients	Frequency %	No. of patients	Frequency %
K	7	26.9	5	12.2
E	6	23.1	11	26.8
C	0	0	2	4.9
D	1	3.8	1	2.4
e	1	3.8	1	2.4
c	2	7.7	1	2.4
NSA*	1	3.8	7	17.1
Kp <sup>a</sup>	1	3.8	1	2.4
Fy <sup>a</sup>	0	0	1	2.4
E+Kp <sup>a</sup>	1	3.8	0	0
K+Kp <sup>a</sup>	0	0	1	2.4
E+K	1	3.8	3	7.3
C+D	2	7.7	2	4.9
C+e	1	3.8	0	0
E+S	0	0	1	2.4
E+Cw	1	3.8	0	0
C+D+Jk <sup>a</sup>	1	3.8	0	0
C+D+E	0	0	1	2.4
C+D+K	0	0	1	2.4
Jk <sup>a</sup> +Fy <sup>a</sup>	0	0	1	2.4
E+Fy <sup>a</sup>	0	0	1	2.4
E+Jk <sup>a</sup>	0	0	1	2.4
<b>Total</b>	<b>26</b>	<b>100</b>	<b>42</b>	<b>100</b>

\* Non-specific antibody



**Figure 1.** Number (%) of specific antibodies in SCD (Figure 1a) & Homozygous Thalassemia Major patients (Figure 1b).

frequencies of the alloantibodies; 30(88%) of the alloantibodies were against the Rh and Kell antigens. The rate of alloimmunization among males was 19% and females 21%.

The rate of alloimmunization in adults (aged 13-32 years) was higher at 14.4% as compared to 4% in children (aged 5-12 years).

*Sickle cell anaemia:* Out of 133 patients, 42(31.5%) developed positive antibody screen in whom 46 IgG alloantibodies were detected (95%CI, 24.87-40.66). Seven patients (16.6%) showed NSA, 23(54.7%) developed one antibody; 10(24%) developed two antibodies, and two

patients had developed three antibodies. The specificities and frequencies of the alloantibodies in Omani patients with SCD are shown in **Table 1** and **Figure 1a**. 38(83%) of the alloantibodies were Rh and Kell antibodies. The rate of alloimmunization among males was 30% and among females 33%.

Furthermore, in the eight patients who had a non-specific antibody, we observed that PG-IAT detected clinically significant antibodies like anti-E, anti-C, anti-D, anti-Jk(a), anti-c, anti-e, anti-s that were masked by an autoantibody in our cohort of multi-transfused patients. PG-IAT was superior in detecting clinically significant allo- antibodies

in the presence of masking autoantibodies as compared to the other techniques employed.

**Discussion.** The factors involved in alloimmunization are complex and includes at least three main contributing elements: the RBC antigenic differences between the blood donor and recipients, the recipient's immune status and the immunomodulatory effect of the allogeneic blood transfusions on the recipient's immune system.<sup>15,16</sup>

This study shows that the prevalence of alloimmunization in Omani homozygous thalassaemia patients was 20% (n=26; 95% CI, 13.9-27.6). Comparing the rate of alloimmunization in Omani thalassaemia patients with that of other populations, it was similar to several countries namely, 16.32% from Iran,<sup>17</sup> and 22% from California<sup>18</sup> and 19% from the CDC data in the USA on Asian and Caucasians patients.<sup>19</sup> But in general, there is a reduction of the frequency of alloimmunization when the patient receives blood from the same ethnic groups like those living in Hong Kong<sup>20</sup> and in Saudi Arabia.<sup>21</sup> The low incidence of immunization found in an old Italian cooperative study of 1984 (5/68;5.4%), could have the same explanation since this study included only thalassaemic patients living in Italy and receiving blood from the same ethnic group.<sup>22</sup> The higher rate of alloimmunization in adults as compared to children is in keeping with other studies where age is a significant factor.<sup>11</sup> The incidence of alloimmunization in transfused Omani patients with SCD was found to be 31.5% (n=42, 95% CI, 24.87-40.66). This rate is similar to that reported in USA, France, Holland, and patients of Asian descent in Brazil.<sup>23-27</sup> It was also noticed that most of the alloantibodies were to Rh and Kell, antigens and that the E antibody had a higher rate.

The Omani population, for historical reasons, is known to be a mixture of more than one ethnic group. So it was expected to detect some antigenic differences among the Omani donors themselves. Also, 10% of donors are non-Omani, so patients receiving blood transfusions will be further exposed to "foreign" antigens. The frequency of alloantibodies may be reduced by limiting the transfusion from donors with the same ethnic origin.<sup>21</sup>

It was noticed that patients with sickle cell anaemia showed a slightly higher rate of alloimmunization (31.5%) than thalassaemia

patients (20%). This datum is consistent with observations by other studies as well. One reason for this observation could be because thalassaemia patients are usually transfused at a younger age and regular intervals. The immune system response will be affected by the patient's age at their first transfusion and number of blood units the patient received.<sup>27</sup> It is believed that transfusions at an early age may offer some immune tolerance and protection against alloimmunization. The relation between the number of blood units transfused and antibody formation is unknown in thalassaemia, but it is a major factor for increased alloimmunization in patients, including SCD, who receive multiple transfusions. However it should be taken into account that SCD is a chronic inflammatory state, and pro-inflammatory stimuli promote alloimmunization.<sup>28,29</sup> Furthermore, age is a significant factor, so children with SCD, who are chronically transfused, might have less inflammation, which could explain their lower rate of alloimmunization.<sup>30,31</sup> However, some antigen-negative patients may not produce antibodies at all or may form only one antibody despite exposure to antigen-positive cells. Studies have suggested at least in SCD patients, that genetic makeup is very relevant to the development of antibodies mainly altered Rh or Kell alleles, and perhaps acquiring these antibodies may be genetically driven.<sup>32,33</sup> In Omani population further studies will be needed to assess the effect of the number of transfusions on the immune response, the effect of the age at which the patient is first transfused, and the genetic makeup of recipients and donors on alloimmunization.

One of the biggest problems in a conventional hospital blood bank is finding the appropriate antigen-negative blood for the allo-immunised patients. Numerous reports show that transfusion of phenotype-matched RBCs (Rh and Kell) can reduce the risk.<sup>19,34</sup> However, there are still few reports revealing that the risk of alloimmunization is still high even when the donor blood is Rh and Kell matched with the recipient.<sup>12,13</sup>

BCSH transfusion guidelines also state that all patients with sickle cell disease and thalassaemia have their full phenotype tested at diagnosis and are given matched blood for C, c, E, e and K.<sup>35,36</sup> Moreover, extended red cell phenotype matching, although useful in preventing the formation of most alloantibodies, may prove impractical to

provide adequate and timely donors for these patients.<sup>37</sup> At present, we too follow this standard recommendation and hope to decrease the rate of alloimmunization. In our hospital it is estimated that the cost of one year of phenotyping for Rh and Kell antigens is about 12,500 OMR (\$32,400) for all the donated units in our blood bank, raising the question whether it is cost effective to phenotype all of these units routinely. Nevertheless, DNA-based phenotyping can overcome certain limitations of serological studies and is beneficial in patients recently transfused or with interfering allo- or autoantibodies.<sup>38</sup>

**Conclusions.** Red cell transfusions are the cornerstone in the management of homozygous thalassaemia major but remain underutilised in SCD patients for fear of complications, although

they can be lifesaving in the context of SCD complications. Nonetheless, they are associated with a considerable risk of alloimmunization as well as iron overload. The current BCSH transfusion guidelines recommend the initial extended red cell phenotyping for both donors and recipients. Thus with careful selection of donated units, coupled with further elaborative studies of the genetic diversity of patients and donor pool will certainly go a long way in reducing the prevalence of red cell alloimmunization. Therefore it will be cost-effective in the long term to choose the appropriate blood donor

**Acknowledgement.** We wish to thank the hospital administration for the use of hospital material for this study.

## References:

- Al-Riyami A, Ebrahim GJ, Genetic blood disorders survey in the Sultanate of Oman. *J Trop Pediatr.* 2003 Jul; 49 suppl 1: i1-20
- Al-Riyami AA, Sulieman AJ, Afifi M, Al-lamki ZM, Daar S, A community- based study of common hereditary blood disorders in Oman. *East Mediterr. Health J* 2001 Nov; 7(6):1004-11
- Alkindi S, Al Zadjali S, Al Madhani A, Daar S, Al Haddabi H, Al Abri Q, Gravell D, Berbar T, Pravin S, Pathare A, Krishnamoorthy R. Forecasting hemoglobinopathy burden through neonatal screening in Omani neonates. *Hemoglobin.* 2010; 34: 135-44. <http://dx.doi.org/10.3109/03630261003677213>
- Wayne AS, Kevy SV, Nathan DG, Transfusion management of sickle cell disease. *Blood* 1993; 81:1109-23
- Hmida S, Mojaat N, Maamar M, Bejaoui M, Mediouni M, Boukef K, Red cell alloantibodies in patients with haemoglobinopathies. *Nouv Rev Fr.hematol* 1994 oct; 36 (5): 363-6.
- Spanos T, Karageorga M, Ladis V, Peristeri J, Hatziliami A, Kattamis C. Red cell alloantibodies in patients with thalassaemia. *Vox Sang* 1990; 58( 1): 50-5. <http://dx.doi.org/10.1111/j.1423-0410.1990.tb02055.x>
- Rosse WF, Gallagher D, Kinney T, Castro O, Dosik H, Moohr J,Wang W, Levy PS, Transfusions and alloimmunization in sickle cell disease. The cooperative study of sickle cell disease. *Blood* 1990 Oct.1; 76(7): 1431-7.
- Aygun B, Padmanabhan S, Paley C, Chandrasekaran V. Clinical significance of RBC alloantibodies and autoantibodies in sickle cell patients who received transfusions. *Transfusion* 2002 Jan; 42(1): 37-43. <http://dx.doi.org/10.1046/j.1537-2995.2002.00007.x>
- Wayne AS, Kevy SV, Nathan DG, Transfusion management of sickle cell disease *Blood*, 1993 Mar 1; 81(5): 1109-23.
- Castellino SM, Combs MR, Zimmerman SA, Issitt PD, Ware RE, Erythrocytes autoantibodies in paediatric patients with sickle cell disease receiving transfusions therapy: frequency, characterization, and significance. *Br. J. Haematol* 1999 Jan; 104(1): 189-94. <http://dx.doi.org/10.1046/j.1365-2141.1999.01127.x>
- Murao M, Viana MB, Risk factors for alloimmunization by patients with sickle cell disease. *Braz J. Med. Biol Res* 2005; 38: 675-682 <http://dx.doi.org/10.1590/s0100-879x2005000500004>
- Chou ST, Jackson T, Vege S, Smith-Whitley K, Friedman DF, Westhoff CM. High prevalence of red blood cell alloimmunization in sickle cell disease despite transfusion from Rh-matched minority donors. *Blood.* 2013;122(6):1062-71. <https://doi.org/10.1182/blood-2013-03-490623>
- Miller ST, Kim H-Y, Weiner DL, Wager CG, Gallagher D, Styles LA, et al. Red blood cell alloimmunization in sickle cell disease: prevalence in 2010. *Transfusion.* 2013;53(4):704-9. <https://doi.org/10.1111/j.1537-2995.2012.03796.x>
- Wayne As, Schoenik SE, Pegelow CH, Financial analysis of chronic transfusions for stroke prevention in sickle cell disease. *Blood* 2000 Oct.1; 96(7):2369-72.
- Alarif L, Castro O, Ofosu M, Dunston G, Scott RB, HLA –B35 is associated with red cell alloimmunization in sickle cell disease. *Clin Immunol Immunopathol* 1986, 38:178-183. [http://dx.doi.org/10.1016/0090-1229\(86\)90136-4](http://dx.doi.org/10.1016/0090-1229(86)90136-4)
- Vichinsky EP, Earles A, Johnson RA, Hoag MS, Williams A, Lubin B, Alloimmunization in sickle cell anemia and transfusion of racially unmatched blood. *N Eng J Med* 1990, 322: 1617-21. <http://dx.doi.org/10.1056/nejm199006073222301>
- Davari K, Soltanpour MS. Study of alloimmunization and autoimmunization in Iranian beta-thalassemia major patients. *Asian J Transfus Sci.* 2016 Jan-Jun;10(1):88-92. <http://dx.doi.org/10.4103/0973-6247.172179>
- Singer ST, Wu V, Mignacca R, Kuypers FA, Morel P, Vichinsky EP Alloimmunization and erythrocyte autoimmunization in transfusion-dependent thalassemia patients of predominantly Asian descent. *Blood.* 2000;96:3369-3373.CrossRefMedlineWeb of Science Google Scholar
- Vichinsky E, Neumayr L, Trimble S, Giardina PJ, Cohen AR, Coates T, Boudreaux J, Neufeld EJ, Kenney K, Grant A, Thompson AA; CDC Thalassaemia Investigators. Transfusion complications in thalassemia patients: a report from the Centers for Disease Control and Prevention (CME). *Transfusion.* 2014 Apr;54(4):972-81; quiz 971. <http://dx.doi.org/10.1111/trf.12348>
- Ho HK, Ha SY, Lam CK, Chan GC, Lee TL, Chiang AK, Lau YL. Alloimmunization in Hong Kong southern Chinese transfusion-dependent thalassemia patients. *Blood.*2001 Jun 15;97(12):3999-4000. PubMed PMID: 11405212
- Abdel Gader AM, Al Ghumlas AK, Al-Momen AM, Transfusion medicine in a developing country-alloantibodies to red blood cells in multi-transfused patients in Saudi Arabia. *Transfus Apher Sci.*2008;39:199-204. <http://dx.doi.org/10.1016/j.transci.2008.09.013>
- Sirchia G, Zanella A, Parravicini A, Morelati F, Rebulla P, Masera G, Red cell alloantibodies in thalassemia major: results of an Italian cooperative study. *Transfusion* 1985 Mar-Apr; 25(2)110-2. <http://dx.doi.org/10.1046/j.1537-2995.1985.25285169198.x>
- Vichinsky EP, Current issues with blood transfusion in sickle cell disease. *Semin. Hematol* 2001 Jan; 38(1 suppl 1): 14-22 <http://dx.doi.org/10.1053/shem.2001.20140>
- Moreira Junior G, Bordin JO, Kuroda A, Kerbaui J, Red cell alloimmunization in sickle cell disease, the influence of racial and antigenic pattern differences between donors and recipients in Brazil. *Am. J Hematol* 1996 Jul; 52(3): 197-200. [http://dx.doi.org/10.1002/\(sici\)1096-8652\(199607\)52:3<197::aid-ajh11>3.0.co;2-d](http://dx.doi.org/10.1002/(sici)1096-8652(199607)52:3<197::aid-ajh11>3.0.co;2-d)

25. Sins JW, Biemond BJ, van den Bersselaar SM, Heijboer H, Rijnveld AW, Cnossen MH, Kerkhoffs JL, van Meurs AH, von Ronnen FB, Zalpuri S, de Rijke YB, Ellen van der Schoot C, de Haas M, van der Bom JG, Fijnvandraat K. Early occurrence of red blood cell alloimmunization in patients with sickle cell disease. *Am J Hematol*. 2016 Aug;91(8):763-9. <http://dx.doi.org/10.1002/ajh.24397>
26. Norol F, Nadiahi J, Bachir D, Desaint C, Guillou Bataille M, Beajeau F, Bierling P, Bonin P, Galacteros F, Duedari N. Transfusion and alloimmunization in sickle cell anemia patients. *Transfus Clin Biol*. 1994; (1): 27-34. [http://dx.doi.org/10.1016/s1246-7820\(05\)80054-0](http://dx.doi.org/10.1016/s1246-7820(05)80054-0)
27. Murao M, Viana MB. Risk factors for alloimmunization by patients with sickle cell disease. *Braz J Med Biol Res* 2005 May; 38(5) : 675-82. <http://dx.doi.org/10.1590/s0100-879x2005000500004>
28. Yu J, Heck S, Yazdanbakhsh K. Prevention of red cell alloimmunization by CD25 regulatory T cells in mouse models. *Am J Hematol*. 2007;82(8):691–696. <https://doi.org/10.1002/ajh.20959>
29. Hendrickson JE, Chadwick TE, Roback JD, Hillyer CD, Zimring JC. Inflammation enhances consumption and presentation of transfused RBC antigens by dendritic cells. *Blood*. 2007;110(7):2736–2743. <https://doi.org/10.1182/blood-2007-03-083105>
30. Murao M, Viana MB. Risk factors for alloimmunization by patients with sickle cell disease. *Braz J Med Biol Res*. 2005;38(5):675–682. <https://doi.org/10.1590/s0100-879x2005000500004>
31. Gill FM, Sleeper LA, Weiner SJ, Brown AK, Bellevue R, Grover R, Pegelow CH, Vichinsky E. Clinical events in the first decade in a cohort of infants with sickle cell disease: Cooperative Study of Sickle Cell Disease. *Blood*. 1995;86(2):776–783.
32. Chou ST, Westhoff CM. Molecular biology of the Rh system: clinical consideration for transfusions in sickle cell disease. *Am Soc Hematol Educ program* 2009: 178-84. <http://dx.doi.org/10.1182/asheducation-2009.1.178>
33. Boturao-Neto E, Chiba AK, Vicari P, Figueiredo MS, Bordin JO. Molecular studies reveal a concordant KEL genotype between patients with hemoglobinopathies and blood donors in Sao Paulo City Brazil. *Haematologica* 2008 sep;93(9): 1408-10. <http://dx.doi.org/10.3324/haematol.12766>
34. Vichinsky EP, Luban NL, Wright E, Olivieri N, Driscoll C, Pegelow CH, Adams RJ. Prospective RBC Phenotype matching in a stroke –prevention trial in sickle cell anemia :a multicenter transfusion trail. *Transfusion* 2001 Sep; 41(9): 1086-92. <http://dx.doi.org/10.1046/j.1537-2995.2001.41091086.x>
35. Guidelines for pre-transfusions compatibility procedures in blood transfusions laboratories-BCSH Blood Transfusions Task Force. *Transfus. Med*. 1996 Sept; 6(3):273-83. <http://dx.doi.org/10.1111/j.1365-3148.1996.tb00079.x>
36. Vichinsky EP, Ohene-Frempong K, Thein SL, Lobo CL, Inati A, Thompson AA, Smith-Whitley K, Kwiatkowski JL, Swerdlow PS, Porter JB, Marks PW. Transfusion and chelation practices in sickle cell disease: a regional perspective. *Pediatr Hematol Oncol* 2011 Mar; 28(2): 124-33. <http://dx.doi.org/10.3109/08880018.2010.505506>
37. Castro O, Sandler G, Houston –Yu P, Rana S, predicting the effect of transfusing only phenotype –matched RBC to patients with sickle cell disease: theoretical and practical implications. *Transfusions* 2002 Jun; 42:684-90. <http://dx.doi.org/10.1046/j.1537-2995.2002.00126.x>
38. Fasano RM, Chou ST. Red Blood Cell Antigen Genotyping for Sickle Cell Disease, Thalassemia, and Other Transfusion Complications. *Transfus Med Rev*. 2016 Oct;30(4):197-201. <http://dx.doi.org/10.1016/j.tmr.2016.05.011>